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Effect of Sulfur Fertilization and Bradyrhizobium Inoculation on Nodulation, Yield and Yield Components of Soybean (*Glycine max* L.) on Nitisols of Jimma Area, Southwestern Ethiopia

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Abstract

A field experiment was conducted at Jimma Agricultural Research Center (JARC) during 2017/18 growing season to study the effect of inoculation of Bradyrhizobium strain and sulfur fertilization on nodulation, grain and straw yields of soybean. The experiment consisted of four levels of S (0, 20, 30 and 40 kg ha⁻¹) and three Bradyrhizobium strains (MAR-1495, SB-12 and Murdock) arranged factorially in completely randomized block design (RCBD) with three replications. Grain and straw yield increased by inoculation of Bradyrhizobium strain whether used alone or in combination with S. Nodulation, parameters (nodule number per plant and nodule dry weight) and number of pods per plant were highly significantly influenced by interaction effect of Bradyrhizobium inoculation strains in combination with sulfur fertilization. The yield and yield components namely (grain yield, hundred seed weight, biomass yield and harvest index) were highly significantly ($P < 0.01$) affected by individual application of sulfur and inoculation of Bradyrhizobium strains alone. Accordingly, the highest grain yield (1496.35 kg ha⁻¹) was obtained from application of S at a rate of 30 kg ha⁻¹ corresponding 30.96% yield advantages compared with control and 1548.55 kg ha⁻¹ grain yield was recorded due to inoculation with Murdock strain corresponding to 30.32% increase over MAR-1495 strain. The results clearly suggested that proper application of S along with Bradyrhizobium strain affect nodulation, grain and straw yield of soybean.

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Keywords

Sulfur, Strain, Nodulation, Grain yield.

Introduction

Soybean (*Glycine max* (L.) Merrill) is one of the most important legume crop cultivated worldwide and domesticated first in China and it is the world's leading source of oil and protein (Hui and Chang, 2010). Depending on the variety, it can grow best from 0-2200m altitude and under rainfall ranging from 300 to 1200mm thus altitude influences temperature that in turn affects the physiological (esp. flowering and maturity)

cycle of the crop. It has the highest protein content (40%) of all food crops and is second only to groundnut in terms of oil content (20%) among food legumes (Gurmu *et al.*, 2009). Soybean requires large amounts of nitrogen (N), phosphorus (P) and potassium (K) as well as a smaller amount of sulfur (S) and some micronutrients. Although soybean requires considerably less P and S than N or K, all are important for plant growth and development. Nitrogen nutrition is not a serious problem as the plant has the inherent ability to obtain most of its

N requirement from the atmosphere through biological fixation by forming a symbiotic relationship with Rhizobium bacteria in the soil. Soybean requires 378 kg ha⁻¹N to complete its growth cycle; however, it has the potential to obtain 60 to 70% of its requirement from biological N fixation (Abendroth and Elmore, 2000). Nonetheless, this relationship could not occur without inoculating the seed with appropriate strain of Bradyrhizobium bacteria.

Ethiopia has suitable natural conditions and vast land for investing soybean production. Southwestern part of Ethiopia, particularly Jimma area is favorable location for soybean cultivation and has an economic importance for smallholders (Wijnands *et al.*, 2011). However, the actual yield productivity of the crop has ranged from 920-1146 kg ha⁻¹, if properly managed; it is possible to achieve the potential yield up to 2000-3500 kgha⁻¹.

Biological nitrogen fixation (BNF) offers an economically attractive and ecologically sound means of improving soybean yield, reducing external N inputs and enhancing the quality of soil, which consequently reduce the dependence on mineral fertilizers that could be costly and unavailable to smallholder farmers when effective and compatible strain inoculated to the soil. Legume crops including soybean obtain between 50 - 80% of their N requirements through biological nitrogen fixation (Solomon *et al.*, 2012). However, Sanginga (2002) reported that the current promiscuous soybean genotypes could not meet all their demand for growth and seed development only by N₂ fixation rather it needs addition of mineral nutrient sources and Bradyrhizobium strain may hardly exist in highly weathered soils because soybean is an exotic crop. Moreover, population of Bradyrhizobias seldom available in soils of deterioration of its quality; therefore, nodulation of soybean may require specific species of Bradyrhizobium for effective N₂ fixation (Abaidoo *et al.*, 2007). However, the success of inoculation depends not only on high quality inoculants and good inoculation practices but also on the establishment of effective and efficient BNF through optimization of the factors that affect its performance such as legume genotype, climatic, edaphic and management factors (Giller, 2001).

Due to low soil fertility status in the country legume crops are generally grown in sever soil conditions, which are inherently low in nutrient content including S and low soil pH especially in Southwestern part of Ethiopia (ATA, 2013). The Ethiopian Soil Information Service is currently involved in mapping the entire country for all

nutrients, and has found extensive areas of S, Zn, and B deficiency (Vanlauwe *et al.*, 2015). Among secondary nutrients S is an essential nutrient for plant growth accounting about 10% of the total N content (Anandham *et al.*, 2007) and legume crops including soybean require it in a similar or more than that of P for high yield and quality (Jemal *et al.*, 2010). On the other hand, in areas like Ethiopia where subsistence farming is common, addition of S through organic decomposition is mostly insufficient to meet the requirement associated with the small yield. Thus, continuous application of N containing fertilizers in the form of DAP excluding sulfur further makes its deficiency worse because of widen N: S ratio. Despite the important roles of S in agriculture, research pertaining to its status in soil and its response in crops are almost nonexistent in Ethiopia (Habtemichael *et al.*, 2007). Therefore, supplying adequate S to the soil has significant potential of increasing the amount of N fixed by legumes and their grain yield, thus improving fertility status of soil (Cazzato *et al.*, 2012). A research finding revealed that lower N accumulation and a yield reduction of legume crops was recorded when S was limiting in the soil (Scherer *et al.*, 2006) and also recognized not only decreased yield but also quality of products when S is deficient (Jemal *et al.*, 2010). Sharma (2014) also reported that S containing amino acids such as methionine and cysteine increased significantly by combined application of N and S and indicated improvement in soybean nutritional value. Therefore, the most important constraints to soybean crop growth may bethose caused by the shortage of plant nutrients (Sand N). Therefore, a field experiment was conducted to evaluate the effect of S fertilization and inoculation of Bradyrhizobium strain on nodulation, yield and yield components of soybean.

Materials and Methods

Description of the study area

A field experiment was conducted at Jimma Agricultural Research Center (JARC) Southwestern Ethiopia under rain fed conditions during 2017/2018 cropping season to investigate the effects of integrated application of S and Bradyrhizobium strain on nodulation and grain yield of soybean. Jimma Agricultural Research Center is located at about 358 km far from Addis Ababa city and 12 km from Jimma town to Southwest direction. Geographically it is located at 07° 40'.212" N latitude and 036°47'.055" E longitude and an altitude of 1763 masl. The average annual maximum and minimum air temperature were 27.2°C and 11.8°C, respectively and the area receives

adequate amount of rainfall, 1198 mm per annum. According to VanRanst *et al.*, (2011), the major reference soil groups in the area dominated by Nitisols, Acrisols, Ferralsols, Vertisols and Planosols.

Treatment Set up and Experimental Procedure

The experiment consist four levels of sulfur (0, 20, 30 and 40 kg ha⁻¹) and three strains of Bradyrhizobium (MAR-1495, SB -12 and Murdock). It was laid out in a completely randomized block design (RCBD) in factorial arrangement with three replications, having a total of 21 treatments accommodating seven rows and net plot area was 3m x 3.9m = 11.7m². Planting was done early June, 2017 based on farmers local planting calendar where two seeds per hill was drilled on ridges made with 0.05 m and 0.60m spacing between plants and each row respectively and covered with soil and thinned to one seed per hill two weeks later, which accommodates a total of plant population levels 333333ha⁻¹. Harvesting was done at physiological maturity by leaving the outer most rows on both sides of each plot to avoid border effects.

Crop Management

The field was prepared on station by removing all unwanted materials and before sowing, the field plowed with oxen, four times to make a fine seed bed. Sulfur was applied as potassium sulfate (K₂SO₄) in solid form. In addition to sulfur, other nutrients, such as starter dose of N fertilizer at 18 kg ha⁻¹ as Urea (Solomon *et al.*, 2012) and P fertilizer as TSP at recommended dose of 23 kg ha⁻¹ was applied during sowing for each treatment. Since S fertilizer was applied in the form of K₂SO₄, the disproportionate addition of potassium in different treatments was counter balanced by the addition of proportionate amount of potassium chloride (KCl).

Soil fertilization and sowing

Soybean seed was selected based on size and healthiness by physical observation. Then the seeds were weighed and surface sterilized by soaking them first with 70% (v/v) ethanol for 10 seconds and 4% (v/v) sodium hypochlorite (NaOCl) solution for five minutes and later washed five times with sterilized water as indicated in Somasegaran and Hoben (1994). Each strain was applied at the rate of 5gm powder inocula per 1kg of seed using the slurry method as described by Woomey *et al.*, (1994). In order to ensure that all the applied inoculum stick to the seed, the required quantity of inoculants was

suspended in 1:1 10% sugar solution. The sugar slurry was gently mixed with dry seed and then with carrier-based inoculant so that all the seeds received a thin coating of the inoculant. Then the strain was mixed thoroughly with seeds. For each inoculation, separate plastic bag was used and care was taken to avoid contact among each strain during sowing. The seed was allowed to shaded air dry for a few minutes and then sown at 60 kg ha⁻¹ seed rate.

Soil sampling and analysis

A composite soil sample was collected using auger in a diagonal pattern at 0-20cm depth before treatment application.

The sample was air-dried, ground using a pestle and mortar and allowed to pass through a 2 mm sieve. Working samples were prepared and analyzed for soil pH, organic carbon (OC), total nitrogen (TN), available phosphorus (Av.P) and cation exchange capacity (CEC) using standard laboratory procedures at Jimma Agricultural Research Center (JARC) soil laboratory. Total N was determined by Kjeldahl digestion, distillation and titration method (Bremner, 1982). Soil pH was determined (1:2.5 soil: water ratio) according to Okalebo *et al.*, (2002). Available P was determined using Bray II extraction method as described by Bray and Kurtz (1945).

Organic carbon content of the soil was determined by reduction of potassium dichromate by OC compound and determined by reduction of potassium dichromate by oxidation-reduction titration with ferrous ammonium sulfate and OC was determined by the method of Nelsen and Sommers (1982). Extracted Phosphorus was determined with atomic absorption spectrophotometer calorimetrically according to molybdenum blue color method described by Murphy and Riley (1962).

Data Collection

Effective number of nodules

It was determined by carefully uprooting and washing the roots of five plants randomly selected from each plot at 50% flowering stage.

The effective nodules were identified by their color where a cross section of an effective nodule made with pocket knife a pink to dark red color whereas the green color indicate non-effective nodules.

Nodule dry weight

It was determined by carefully uprooting and detached the nodules from the root and dried in oven dry until constant weight is attained then the dried nodule was weighed with sensitive balance.

Yield and Yield components

Number of pods per plant

It was counted from five randomly selected plants of the middle rows at the time of harvesting from each plot by visual observation and their averages were recorded.

Number of seeds per pod

Was determined as the total number of seeds counted per pod in each plot during harvesting from five randomly selected plants.

Grain yield (kg ha⁻¹)

It was measured from each plot and converted into hectare base.

Biomass yield (kg ha⁻¹)

Plants from the net plot area was harvested at physiological maturity, after gained constant weight the dried straw was weighed and converted into hectare base.

Hundred seed weight (gm)

Was determined by counting seeds manually from ten randomly selected plants and its weight was measured by sensitive balance and adjusted to 10% standard moisture content of legumes. Finally, the average value was taken as hundred seed weight

Harvest Index (%)

It was expressed as the ratio of economic yield per plant to the total above ground biomass calculated as follow;

$$HI(\%) = \frac{GY \text{ kg/ ha}}{TBY \text{ kg/ ha}} \times 100$$

Where, HI= harvest index, GY=Grain yield (at 10% moisture level), TBY=Total biomass yield.

Statistical Analysis

The collected data were summarized and statistically analyzed using the analysis of variance (ANOVA) procedure for RCBD using SAS 9.3 version software (SAS, 2012). Treatment means that differed significantly was separated using LSD procedure at 5% level of significance. Correlation coefficient was determined for parameters using the same software to determine relations between yield and yield contributing characters.

Results and Discussion

Number of effective nodules

The ANOVA table showed that there was highly significant interaction effect on number of effective nodules per plant ($P < 0.01$) due to combined application of sulfur and bioinoculant strain. Accordingly, the highest effective nodule number per plant (15.33) having pink in color was recorded from application of S at rate of 20 kg ha⁻¹ in combination with MAR-1495 strain which produced 31.70% nodule number advantages compared with control (no S fertilization) combined with SB-12 strain. The increase in nodule number is an indication that native strains of Bradyrhizobium strain bacteria could form symbiotic relation with soybean are absent in the soils of study site. Effective strains are causes larger few nodules congregated on main roots and are pink and/ red in colour due to the presence of iron containing substance called leg hemoglobin while the nodules produced by ineffective strains have neither leg hemoglobin nor pink or red pigmentation. This result is supported by (Danso *et al.*, 1987 and Wu and Arima, 1992) who reported varietal differences in terms of nodule numbers was observed due to inoculation of different rhizobial strains and observations in Chinese milk vetch, respectively.

Moreover, legumes have a high internal P requirement for their symbiotic N₂ fixation. Singleton *et al.*, (1985) reported that, in addition to the nodule formation, deficiency of P in legumes also markedly affects the development of effective nodules and the nodule leg hemoglobin content.

It is therefore, suggested that the presence of high amount of available phosphorous in soils as in the soils of the experimental field may be beneficial to nodule nitrogen fixation through the prevention of the decrease of the phosphorous concentration in the plants at the later growth stage.

Nodule dry weight

Nodule dry weight was significantly ($P < 0.05$) affected due to the interaction of sulfur fertilization in combination with inoculation of rhizobial strain (Table 3). The maximum nodule dry weight ($4.57 \text{ gm plant}^{-1}$) was obtained from application of sulfur at 20 kg ha^{-1} in combination with MAR-1495 strain, which is statistically at par with inoculation of Murdock strain in combination with application of S at 30 kg ha^{-1} ($4.50 \text{ gm plant}^{-1}$), while the minimum nodule dry weight ($2.63 \text{ gm plant}^{-1}$) was recorded from application of 40 kg ha^{-1} S in combination with SB-12 strain. The extent of establishment of inoculated rhizobia in soil through biofertilizer is dependent on the type of legumes, effectiveness of strains, inorganic content of N, level of available P, K and presence of compatible forms of secondary nutrients (esp. S). At field condition, soybean has shown positive correlation in yield with nodule weight not with nodule number. Plant nodulated by ineffective strains showed lower dry matter yield, grain yield and N content, irrespective of the number of nodules than those nodulated by effective rhizobium strains. The yield increase obtained from the use of biofertilizer relates not only to the N enrichment but also to the antagonistic effect on harmful organisms in the rhizosphere. This is in agreement with the report of Lindermann and Ham (1979) who observed that nodule dry weight was significantly dependent on Rhizobium strain and soybean variety interaction.

Number of pods per plant

The ANOVA table showed that there was highly significant interaction effects ($P < 0.01$) on number of pods per plant due to application of S in combination with Bradyrhizobium. Sulfur fertilization at a rate of 30 kg ha^{-1} in combination with Murdock strain recorded significantly the highest number of pods per plant (52.60) which produced 32.07% more number of pods per plant as compared to inoculation of MAR-1495 strain alone. This revealed the presence of positive interactive effect of S and Bradyrhizobium strain on number of pods per plant on nitisols of Jimma area. Application of S in combination with Bradyrhizobium strains plays an important role in physiological and developmental processes in plant life and the favorable effect of these important nutrients combination (N and S) might accelerate the growth processes, which ultimately resulted in increased yield and quality. The number of pods per plant formed is influenced by the treatment given by Bradyrhizobium strain. This proves that

bacteria that work with root nodules affect plants in forming pods. This result agreed with Jumrawati (2010) who reported that number of pods produced by soybean is largely determined by vegetative growth such as photosynthesis rate and assimilation.

Grain yield

The ANOVA table showed that there was non-significant interaction effect due to combined application of S and bio-inoculant but main effect was observed highly significantly ($P < 0.01$) affected due to application of S and Bradyrhizobium inoculation separately. Accordingly, the maximum yield ($1496.35 \text{ kg ha}^{-1}$) was obtained from application of S at a rate of 30 kg ha^{-1} , which produced 30.95% yield advantages compared with control. In case of Bradyrhizobium inoculation $1548.55 \text{ kg ha}^{-1}$ was recorded from inoculation of Murdock strain that accounts 30.32% and 22.64% yield advantages compared with MAR-1495 and SB-12 strains, respectively. From this, it has demonstrated that integration of sulfur with Bradyrhizobium inoculation did not increase grain yield compared to the individual application. The result also revealed that grain yield of inoculated plants showed the increasing trends as the rates of S. This implies that S application increases the effectiveness of residual indigenous rhizobia rather than the inoculated isolate. It is not clear how high rates of S application reduce the effectiveness of inoculated isolate. However, there was a main effect on grain yield due to S fertilization. This result agreed with the finding of (Kuykendall *et al.*, 1996) who reported that maximum seed yield have been recorded by inoculation of soil with biological strains.

Application of S plays an important role in physiological and developmental processes in plant life and the favorable effect of these important nutrients combination (N and S) might accelerate the growth processes, which ultimately resulted in increased grain yield and quality of the crop. The synergistic effect of N and S is common thus, might be due to utilization of high quantities of nutrients through their well-developed root system and nodules, which might have resulted in better growth and yield. Sulfur and N relationship was established in many studies in terms of dry matter and yield in several crops. Barney and Bush (1985), while working on tobacco plant concluded that there was apparent accumulation of one nutrient when the other nutrient was limited and the accumulated nutrient was used in protein synthesis when the treatment were reversed. A shortage in S supply to the crops lowers the utilization of available soil N,

thereby increasing nitrate leaching thus N status in soil become unsatisfactory. This result agreed with the findings of Hussain *et al.*, (2011).

Application of S might have increased the availability of nutrient to soybean plant due to improved nutritional environment, which in turn, favorably influenced the energy transformation activation of enzymes, chlorophyll synthesis as well as increased carbohydrate metabolism (Dhage *et al.*, 2014). It also constitutes the main element of amino acids including cysteine and methionine, which are of essential nutrient value and can increase grain yield. Therefore, soybean production and protein quality could significantly improve by increasing the concentration of the S containing amino acids through S fertilization. Thus, yield and quality of legume seeds including soybean are limited by the amount of S partitioned to the seeds as reported by (Tan *et al.*, 2010) that supports the current result.

Biomass Yield

The ANOVA table showed that there was non-significant interaction effect on biomass yield due to combined application of S and bio-inoculation (Table 6). However, the main effect of S fertilization and inoculation of strain separately was highly significant ($P < 0.01$) effects on biomass yield, which might be the presence of indigenous strain in the soil and less competent capacity of the inoculated strain. Accordingly, the highest biomass yield ($4924.10 \text{ kg ha}^{-1}$) was recorded from S fertilization at a rate of 30 kg ha^{-1} , which is statically at par with plots treated with 40 kg ha^{-1} S (4825 kg ha^{-1}). Application of 30 kg ha^{-1} S produced 8.24 % biomass yield advantages over control. This showed that S is an essential nutrient for plant growth accounting 10% of the total N content (Anandham *et al.*, 2007) and legume crops including soybean generally require S in a similar quantity or more than that of P for high yield and quality there by maximize above ground biomass, which in line with (Jemal *et al.*, 2010).

With regard to strain, the maximum biomass yield ($1548.55 \text{ kg ha}^{-1}$) was obtained from inoculation of Murdock strain while the minimum biomass yield ($1079.02 \text{ kg ha}^{-1}$) was recorded from inoculation of SB-12 strain. From the current result, inoculation of Murdock strain produced 22.64% and 30.32% biomass yield advantages compared with SB-12 and MAR-1495 strain, respectively. Inoculation of Bradyrhizobium strain to soybean seed increased growth and yield contributing parameters including seed weight. Each strain has its

own synergetic effect on production of seed weight as seed weight also increased when rhizobium strains (significantly greater for Murdock) were applied individually but not in combination with S.

An increase in biomass was observed because rhizobial inoculation is known to increase yield of several legume crops by increasing nodulation and biomass of root and shoot, which is supported by (Subba, 1986).

Hundred Seed Weight

The ANOVA table showed that hundred seed weight was non-significant interaction effects due to combined application of S and Bradyrhizobium strain but significantly ($P < 0.05$) affected by main effects due to S application alone. The highest hundred seed weight (11.32 gm) was recorded from application of S at a rate of 20 kg ha^{-1} , which produced 10.60% advantages compared with control plots. This result showed soybean seed quality can significantly improve by increasing the concentration of S containing nutrient sources because S application is an important nutritional element to get full and better soy bean seed. The role of S in seed production of soybean has also reported by (Jamal *et al.*, 2005 and Dhage *et al.*, 2014) who found that soybean seed yield and straw yield increased significantly due to application of 60 kg ha^{-1} S followed by 40 kg ha^{-1} S over control.

Harvest Index

The ANOVA table showed that harvest index was observed to be significantly ($P < 0.05$) affected by inoculation of strain alone and the interaction effect of fertilizer and strain was highly affected at ($P < 0.01$). Sulfur is an important nutrient for plant growth and development. Sulfur interaction with N are directly related to the alteration of physiological and biochemical responses of crops, and thus required to be studied in depth. This would help to understand nutritional behavior of S in relation to N and provide guidelines for inventing balanced fertilizer recommendations in order to optimize yield and quality of crops. A strong interaction of S and N for seed yield was found in Soybean (Jamal *et al.*, 2006b).

Correlation Analysis

Correlation analysis between growth and yield related parameters is presented in (Table 4). Crop yield is a cumulative interaction effect of all the dependent and

independent characters in an experiment. This is because it has a positive relationship with those parameters and contributes a great growth performances and yield parameter for the plants. According to present result, grain yield was highly significantly ($P < 0.01$) and positively correlated with number of pods per plant ($r=0.52$), and harvest index ($r = 0.53$) and significantly correlated with biomass yield ($r = 0.38$) indicating they have direct relationship on yield. Number of pods per

plant was highly significantly ($P < 0.01$) and positively correlated with biomass yield ($r=0.44$) and significantly ($P < 0.05$) correlated with nodule dry weight ($r=0.35$). Number of pods per plant was significantly ($P < 0.05$) and positively correlated with harvest index ($r=0.35$). Number of nodule and nodule dry weight was significantly ($P < 0.05$) and positively correlated with hundred seed weight $r=0.32$ and $r = 0.37$, respectively.

Table.1 Soil Physico - Chemical properties of experimental site before sowing in 2017/18 season

Soil Properties	Value	Rating	References
pH (1:2.5 H ₂ O)	5.52	Moderate acidic	Tekalign Mamo (1991)
Soil BD (g cm ⁻³)	1.25	Optimum	Hunt and Gilkes (1992)
CEC (cmol(+) Kg ⁻¹)	16.06	Medium	Hazelton and Murphy (2016)
Total N (%)	0.10	Low	Tekalign Mamo (1991)
Avail. P (mg kg ⁻¹)	8.18	Low	Landon (1991)
OC (%)	1.84	Medium	Charman and Roper (2007)
OM (%)	3.17	Medium	Nelson and Sommers (1982)
Soil Textural Class	Clay loam	Ideal	Onwueme and Sinha (1991)

Table.2 Interaction effect of S and Bradyrhizobium strains on soybean nodule number, nodule dry weight and number of pods plant⁻¹

Treatments Bradyrhizobium Inoculation	Sulfur rates (kg ha ⁻¹)			
	0	20	30	40
	Number of nodule plant⁻¹			
MAR-1495 strain	11.27e	15.33a	13.47bc	14.33abc
SB -12 strain	10.47e	13.07cd	13.20bcd	13.33bcd
Murdock strain	13.60bc	14.80ab	13.53bc	11.73de
LSD (0.05)	1.64			
CV (%)	6.89			
	Nodule dry weight (gm)			
MAR-1495 strain	3.53a-d	4.56a	4.32ab	3.17cd
SB -12 strain	2.68d	4.03abc	4.03abc	2.63d
Murdock strain	3.35bcd	3.27cd	4.50a	3.93abc
LSD (0.05)	1.05			
CV (%)	15.01			
	Number of pods plant⁻¹			
MAR-1495 strain	35.73e	38.80de	43.40cd	43.20cd
SB -12 strain	43.53cd	43.87cd	42.13cde	38.73de
Murdock strain	44.47bcd	51.20ab	52.60a	46.87abc
LSD (0.05)	7.15			
CV (%)	7.41			

Different small letters denote significant difference between treatments.

Table.3 Effects of sulfur and Bradyrhizobium on soybean yield, biomass, hundred seed weight and harvest index at Jimma in 2017/18 cropping season

Treatments	Grain yield (Kg ha ⁻¹)	Biomass yield (Kg ha ⁻¹)	100- Seed weight (gm)	Harvest Index(%)
Sulfur rate (kg ha⁻¹)				
0	1033.15c	4518.50b	10.12b	21.93
20	1243.93b	4549.10b	11.32a	21.96
30	14963.35a	4924.10a	11.03a	22.21
40	1327.37b	4825.90a	11.18a	21.30
LSD (0.05)	154.69	256.76	0.79	ns
P- value	**	**	*	-
Bradyrhizobium inoculation				
MAR-1495	1079.02b	4809.00a	11.20	20.64c
SB-12	1198.02b	4441.70b	10.75	21.94b
Murdock	1548.55a	4862.50a	10.80	22.96a
LSD (0.05)	133.97	222.36	ns	0.96
P - value	**	**	-	**

Different small letters denote significant difference between treatments.

Table.4 Pearson¹ Correlation Coefficients between growth and yield component of soybean

Variables	GY	BY	NPP	NN	NDW	HSW	HI
GY	1.00						
BY	0.38*	1.00					
NPP	0.52**	0.44**	1.00				
NN	0.12 ^{ns}	0.25 ^{ns}	0.14 ^{ns}	1.00			
NDW	0.6 ^{ns}	0.35*	0.26 ^{ns}	0.27 ^{ns}	1.00		
HSW	0.05 ^{ns}	0.12 ^{ns}	-0.05 ^{ns}	0.32*	0.37*	1.00	
HI	0.53**	-0.17 ^{ns}	0.35*	-0.14 ^{ns}	0.04 ^{ns}	-0.04 ^{ns}	1.00

Where, GY = Grain yield, BY = Biomass yield, NPP = Number pods per plant, NN = Number of nodules per plant, NDW= Nodule dry weight, HSW= Hundred seed weight, HI = Harvest index, ** = significant at P < 0.01, * = significant at P < 0.05, ns = non-significant

Currently increasing soybean production is a mandatory practice through integrated soil fertility management because it has greater prospects in improving soil fertility status and achieving high crop yield via combination of mineral fertilizers and bio-inoculants; which could synergistically improves the overall properties of the soil. Based on the result it is possible to conclude that application of sulfur in combination with Bradyrhizobium inoculants is an imperative to enhance not only soybean production but also sustainable health of soil. Thus, efforts should be made, wherever possible, to introduce inoculation technology to the farming community in Jimma area. Sulfur application at the rate of 20kg ha⁻¹ and 30kg ha⁻¹ in combination with Murdock Bradyrhizobium strain increased nodulation and yield of soybean. More research based on standard methods needs to be undertaken to assess the contribution of

nitrogen-fixing plants to the overall nitrogen budget. The use of rhizobial inoculant is at its infancy in Ethiopia, and so far, it is not one of the research priorities of Agricultural Research Institutions in the country. Rhizobial inoculants are not locally available and farmers are not aware about this new technology. Therefore, more efforts need to be done to popularize this cheap and eco-friendly technology among resource poor farming community of the nation. For an alternative use, Murdock and MAR-1495 strains can recommended for soybean inoculum in Jimma area.

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Data availability statement

The data that support the findings of this study are available from the corresponding author upon reasonable request.

References

1. Abaidoo R. C., Keyser H. H., Singleton P. W., Dashiell, K. E. and Sanginga, N. (2007). Population size distribution and symbiotic characteristics of indigenous *Bradyrhizobium* spp. that nodulate TGx soybean genotypes in Africa. *App. Soil Ecol.* 35: 57 - 67.
2. Agricultural Transformation Agency (ATA) (2013). Status of soil resources in Ethiopia and priorities for sustainable management. GSP for Eastern and Southern Africa. Mar 25-27, 2013, Nairobi, Kenya.
3. Anandham R, Sridarb R, Nalayinic P, Poonguzhalia S, Madhaiyana M, Tongmin SAA (2007) Potential for plant growth promotion in groundnut (*Arachis hypogaea* L.) cv. ALR-2 by co-inoculation of sulfur-oxidizing bacteria and *Rhizobium*. *Microbiol. Res.* 162:139-153.
4. Barney Jr PE, Bush LP (1985) Interaction of nitrate and sulphate reduction in tobacco. I. influence of availability of nitrate and sulphate. *J Plant Nutr* 8: 507-515
5. Bray R. H. and L. T. Kurtz. 1945. Determination of total organic and available forms of phosphorous in soils. *Journal of Soil Science*, 63: 370 – 379.
6. Bremner J M, Mulvaney C S. Total Nitrogen In: AL Page, RH Miller, DR Keeney (Eds.) *Methods of soil analysis. Part 2, Agronomy.* 1982; 9:595-624.
7. Cazzato E, Laudadio V, Stellacci A M, Ceci E, Tufarelli V (2012). Influence of S application on protein quality, fatty acid composition and nitrogen fixation of white lupin (*Lupinus albus* L.). *Eur. Food. Res. Technol.* 235:963-969.
8. CDI (Center for Development Innovation), WUR (Wageningen University and Research). 2011. Developing new value-chains for soybean in Ethiopia. Danso, C. Hera, and C. Douka, "Nitrogen fixation in soybean as influenced by cultivar and *Rhizobium* strain," *Plant and Soil*, vol. 99, no. 1, pp. 163–174, 1987.
9. Dhage S J, Patil V D, Patange M J (2014). Effect of various levels of phosphorus and sulfur on yield, plant nutrient content, uptake and availability of nutrients at harvest stages of soybean *Glycine max* (L.). *Int. J. Curr. Microbiol. Appl. Sci.* 3(12):833-844.
10. Giller K. E. (2001). Nitrogen fixation in tropical cropping systems. 2nd edn. Wallingford, United Kingdom, CAB International. 423 pp.
11. Gurmu F., Mohammed, H. and Alemaw G., 2009. Genotype x environment interactions and stability of soybean for grain yield and nutrition quality. *African Crop Science Journal*, 17(2).
12. Habtemichial K H, Singh B R, Aune J B (2007). Wheat response to N₂ fixed by Fababebean (*Vicia faba* L.) as affected by sulfur fertilization and rhizobial inoculation in semi-arid Northern Ethiopia. *J. Plant Nutr. Soil Sci.* 170:412-418.
13. Hussain K, Islam M, Siddique M T, Hayat R, Mohsan S (2011). Soybean growth and nitrogen fixation as affected by sulfur fertilization and inoculation under rain fed conditions in Pakistan. *Int. J. Agric. Biol.* 13:951-955.
14. Jamal A, Fazli SI, Ahmad S, Abdin M Z, Yun S J (2005). Effect of Sulfur and Nitrogen application on Growth Characteristics, Seed and Oil Yields of Soybean Cultivars. *Korean J. Crop Sci.* 50(5):340-345.
15. Jamal A, Fazli S I, Ahmad S, Abdin M Z, Yun S J (2005). Effect of Sulfur and Nitrogen application on Growth Characteristics, Seed and Oil Yields of Soybean Cultivars. *Korean J. Crop Sci.* 50(5):340-345.
16. Jamal A, Moon YS, Abdin MZ (2010). Enzyme activity assessment of peanut (*Arachis hypogea* L.) under slow-release S fertilization. *Aust. J. Crop Sci.* 4(3):169-174
17. Kuykendall, F. M. Hashem, and W. J. Hunter, "Enhanced competitiveness of a *Bradyrhizobium japonicum* mutant strain improved for nodulation and nitrogen fixation," *Plant and Soil*, vol. 186, no. 1, pp. 121–125, 1996.
18. Lindermann and G. E. Ham, "Soybean plant growth, nodulation, and nitrogen fixation as affected by root temperature," *Soil Science Society of America Journal*, vol. 43, no. 6, pp.1134–1140, 1979.
19. Murphy J, Rilly J. A modified single solution method for the determination of phosphate in natural waters. *Anal chim. Acta.* 1962;27:31-36.
20. Nelson, D. W. and L. E. Sommers, 1982 Total carbon, organic carbon, and organic matter. *Methods of soil analysis part 3-chemical methods, (methods of soil an 3)*, pp. 961-1010.

21. Okalebo J R, Gathua K W, Woormer P L. Laboratory methods of soil and plant analysis: A working manual. 2nd Edition. TSBF-CIAT 2002; 128.
22. Sanginga, N., Okogun, J. A., Vanlauwe, B. and Dashiell, K (2002). The contribution of nitrogen by promiscuous soybeans to maize-based cropping in the moist savanna of Nigeria. *Plant and Soil* 241: 223 - 231.
23. Scherer H W, Pacyna S, Spoth K, Schulz M (2006). Sulfur supply to peas (*Pisum sativum* L.) influences symbiotic N₂ fixation. *Plant Soil Environ.* 52(2):72-77.
24. Singleton, H. M. Abdelmagid, and J. W. Tavares, "Effect of phosphorus on the effectiveness of strains of *Rhizobium japonicum*," *Soil Science Society of America Journal*, vol. 49, no. 3, pp. 613–616, 1985
25. Solomon T, Pant L M, Angaw T (2012). Effects of Inoculation by *Bradyrhizobium japonicum* Strains on Nodulation, Nitrogen Fixation, and Yield of Soybean (*Glycine max* L.) Varieties on Nitisols of Bako, Western Ethiopia. *ISRN Agronomy* (261475):1-8.
26. Solomon, T., Lalit, M. P., and Tsige, A. (2012). Effects of inoculation by *Bradyrhizobium* strains on nodulation, nitrogen fixation and yield of soybean (*Glycine max* L. Merrill) varieties on Nitisols of Bako, Western Ethiopia. Department of natural resource management, college of agric and env'tal science, Haramaya University. International scholarly research network.
27. Somasegaran P, Hoben H J (1994). Handbook for Rhizobia—Methods in Legume-Rhizobium Technology. Springer-Verlag, Heidelberg, Germany. pp. 10-52.
28. Subba Rao, K. V. B. R. Tilak and C. S. Singh, "Dual inoculation with *Rhizobium* sp. and *Glomus fasciculatum* enhances nodulation, yield and nitrogen fixation in chickpea (*Cicer arietinum* Linn.)," *Plant and Soil*, vol. 95, no. 3, pp. 351–359, 1986.
29. Tan Q, Zhang L, Grant J, Cooper P, Tegeder M (2010). Increased Phloem Transport of S-Methyl methionine Positively Affects Sulfur and Nitrogen Metabolism and Seed Development in Pea Plants. *Plant Physiol.* 15:1886-1896.
30. Vanlauwe B, Descheemaeker K, Giller K E, Huising J, Merckx R, Nziguheba G, Wendt J, Zingore S (2015). Integrated soil fertility management in sub-Saharan Africa: unraveling local adaptation. *Soil* 1:491-508
31. Wijnands, J. H. M., Gurmesa, N. D., Lute, J. C. M. and Van Loo, E. N., 2011. *Ethiopian soya bean and sunflower value chains: Opportunities and challenges* (No. 2011-016). LEI, part of Wageningen UR.
32. Wu and Y. Arima, "Effect of *Rhizobium* inoculation and application of N,P,K fertilizer on the growth and nitrogen fixation of field-grown Chinese milk vetch," *Soil Science and Plant Nutrition*, vol. 38, no. 1, pp. 75–84, 1992.

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